SESQUITERPENE LACTONES FROM A POPULATION OF HELENIUM INTEGRIFOLIUM¹

Alfonso Romo de Vivar,* Guillermo Delgado, Manuel Soriano-García, Rubén A. Toscano, Eduardo Huerta, and Rosa G. Reza-Garduño

Instituto de Química de la Universidad Nacional Autónoma de México, Ciudad Universitaria, Circuito Exterior, Coyoacán 04510, México, D.F.

Previous publications (1-7) have described some of our work on the chemical constituents of several *Helenium* species (Compositae), a genus of ca. 50 members (8) located mainly on the American continent. The biological and agricultural importance of this group of plants is well known inasmuch as several species are highly toxic and, when eaten by dairy cattle, impart a bitter taste to their milk (9). Helenanolides are the main constituents of this genus, although its classification by flavonoid chemistry has been proposed (10, 11).

Here we report that the sesquiterpene lactones carabrone [1] (12) and linifolin A [2] (13, 14) were isolated from the extracts of aerial parts of a population of *Helenium integrifolium* (H.B.K.) Benth. & Hook. [Dougaldia integrifolia (H.B.K.) Cass.].



Because of the various stereochemical arrangements and conformational distortions of the cycloheptane ring, and Xray analysis of linifolin A [2] was undertaken. Crystal data for 2 are listed in the Experimental Section. Figure 1 shows the computer generated drawings of two crystallographically independent molecules (A and B) existing in the solid state and differing only slightly in the conformation of the seven-membered ring.

Table 1 lists atomic coordinates of the conformer A, and tables listing atomic coordinates, bond lengths, bond angles, hydrogen coordinates, and structure factors of both conformers (A and B) are available on request to the senior author.

The conformation of the cycloheptane ring of **2** approximates to a half-distorted chair, similar to the conformations found in stramonin B (15) and bromohelenalin (16), but different from the conformations found in florigrandin (17), rudmollin (18), and hymenolane (19), where the cycloheptane ring is a boat. Figure 1 also indicates that the observed values for $J_{6,7}$ and $J_{7,8}$ (4.7 and 9.2 Hz) (13) accord with structure **2** where the dihedral angles involving H(6)-H(7) and H(7)-H(8) are 32 and



163°, respectively. The cyclopentane ring is an envelope with C(5) as the flap.

EXPERIMENTAL

PLANT MATERIAL.—Aerial parts of *H. integrifolium* were collected near Nevado de Toluca volcano, State of Mexico, in September 1983. Reference specimens are deposited in the National Herbarium, Instituto de Biología de la Universidad Nacional Autónoma de México (voucher GD 1153).

EXTRACTION AND ISOLATION.—Air-dried aerial pars (2.58 kg) were twice extracted with

¹Contribution No. 843 of Instituto de Química de la UNAM.



FIGURE 1. Computer-generated drawings of two crystallographically independent molecules (A and B) of linifolin A [2].

hexane to afford 37.1 g residue, and then with Me₂CO-CH₂Cl₂ (1:1, twice) affording 49.1 g syrup. The hexane extract was charged on a silica gel column eluted with hexane-EtOAc gradient beginning with 100% hexane. Carabrone [1] [mp 86-88°, lit. 90-91° (12)] was obtained from fractions eluted with hexane-EtOAc (4:1). The Me₂CO-CH₂Cl₂ extract was also chromatographed on a silica gel column using a hexane-Me₂CO gradient elution system. Compound 1 was also isolated from the less polar fractions of this chromatography, and linifolin A [2] was crystallized from the fractions eluted with hexane-Me₂CO (7:3) (mp and mmp 188-190°). The total yield of 1 was 0.15% and of 2, 0.07% (dry wt). The compounds were identified by comparison of spectral data (ir, uv, ¹H and ¹³C nmr) and by direct comparison with authentic samples.

CRYSTALLOGRAPHIC ANALYSIS OF 2 .--- X-Ray structure analysis .--- Linifolin A [2] was crystallized by slow evaporation from hexane/Me2CO solution. Crystal data: $C_{34}H_{40}O_{10}$, $M_w = 608.7$, orthorhombic, space group $P2_12_12_1$, a= 6.463(5), b=12.872(4), c=37.911(13)Å, V=3154(1)Å³, D_c=1.28 g cm⁻³, F(000)=1296, $T=293 \text{ K}, Z=8, \mu(MoK\alpha)=0.88 \text{ cm}^{-1}$. Using a crystal 0.32×0.35×0.48 mm, lattice parameters were obtained from 25 machine-centered reflections with $4.3 < 2\theta < 21.5^{\circ}$. Reflections in one octant of reciprocal space were measured with an index range of h $0 \rightarrow 6$, $\ell 0 \rightarrow 13$, $\ell 0 \rightarrow 40$, using the w-scanning mode with graphite-monochromatized MoKa radiation on a Nicolet R3m four-circle diffractometer, variable scan speed, scan width 1.0 (θ°), and two standard reflections $(0, 2, \overline{9}; 0, 4, 1)$ monitored every 50 measure-

TABLE 1.	Positional Parameters ($\times 10^4$) and Equivalent Isotropic Temperature
	Factors (Å ²) for conformer A of linifolin A [2]
	$U_{eq} = (U_{11} \times U_{22} \times U_{33})^{1/3}$

Atom	x	у	z	U _{eq}
O(1a)	12402(7)	9877(3)	2575(1)	67(1)
O(2a)	9047(6)	9167(3)	1948(1)	48(1)
O(3a)	4678(6)	6963(3)	2008(1)	53(1)
O(4a)	5218(8)	6152(2)	1493(1)	82(2)
O(5a)	12307(8)	9118(4)	1735(1)	94(2)
C(1a)	8479(8)	8134(4)	2875(1)	39(2)
C(2a)	9694(11)	8505(5)	3196(1)	56(2)
C(3a)	11178(11)	9167(5)	3115(1)	61(2)
C(4a)	11152(9)	9382(4)	2737(1)	49(2)
C(5a)	9147(9)	8911(3)	2588(1)	39(2)
C(6a)	9483(8)	8417(3)	2218(1)	34(1)
C(7a)	8098(8)	7471(4)	2156(1)	39(2)
C(8a)	5800(9)	7629(4)	2245(1)	41(2)
C(9a)	5234(9)	7312(4)	2619(1)	46(2)
C(10a)	6129(10)	7945(4)	2921(1)	44(2)
C(11a)	8035(10)	7045(4)	1792(1)	50(2)
C(12a)	5877(11)	6664(4)	1734(1)	57(2)
C(13a)	9481(12)	6946(5)	1548(2)	73(3)
C(14a)	5643(12)	7396(5)	3266(2)	70(2)
C(15a)	7673(10)	9879(4)	2576(2)	53(2)
C(16a)	10585(12)	9437(5)	1720(2)	68(2)
C(17a)	9752(16)	10188(6)	1454(2)	107(4)

ments. The intensities were corrected for Lorentz and polarization effects, but no absorption corrections were applied. Of the 2389 reflections within the 20 range of 3°-45° collected, 2132 had values of |Fo|² that were greater than three times their estimated standard deviations, and these were used in the final refinement of structural parameters. The structure was solved by direct methods (20) with some difficulties, which are inherent to the space group. The correct structure was found using 200 phases with |E|>1.4 and ten reflections in the starting set, and refined by cascade matrix least-squares techniques with anisotropic temperature factors for the nonhydrogen atoms. The H atoms of the tertiary CH, secondary CH₂, and primary CH₃ groups were assigned coordinates based on the expected bonding geometry. All H atom coordinates were refined and had a fixed isotropic temperature factor, $U=0.06 \text{ Å}^2$. The function minimized was $\Sigma \omega (\Delta F)^2$ with a statistical weight of the form $\omega = \sigma^2(F_0) + 0.003$ (Fo)^{2 -1}, where σ is the standard deviation of the observed amplitudes based on counting statistics. The final conventional R factor was 0.053 and $\omega R = 0.076 \ (\omega R = [\Sigma \omega (|F_0| - |F_0|)^2 / \Sigma |F_0|^2]^{1/2});$ scattering factors for O, C, and H atoms were from International Tables for X-ray Crystallography (21), and the isotropic extinction parameter X=0.0004. A final difference Fourier synthesis revealed no peaks higher than those assigned earlier to hydrogen atoms, and the weighting appeared reasonable.

The final atomic coordinates for the nonhydrogen atoms of the conformer A are listed in Table 1, and pertinent data of the crystallographic analysis are available on request to the senior author.²

ACKNOWLEDGMENTS

We thank Prof. José Luis Villaseñor, National Herbarium, Instituto de Biología de la Universidad Nacional Autónoma de México, for the identification of the plant.

This work was supported in part by the Consejo Nacional de Ciencia y Tecnología (CON-ACYT), México.

LITERATURE CITED

- A. Romo de Vivar and J. Romo, *Chem. and Ind.*, 882 (1959).
- A. Romo de Vivar and J. Romo, Ciencia (Méx.), 21, 33 (1961).

- W. Herz, A. Romo de Vivar, J. Romo, and N. Viswanathan, J. Am. Chem. Soc., 86, 19 (1963).
- W. Herz, A. Romo de Vivar, J. Romo, and N. Viswanathan, *Tetrabedron*, **19**, 1359 (1963).
- A.T. Ozawa, P.A. Rivera, and A. Romo de Vivar, *Rev. Latinoam. Quím.*, 14, 40 (1983).
- 6. A. Romo de Vivar and G. Delgado, Tetrahedron Lett., 579 (1985).
- 7. A. Romo de Vivar, G. Delgado, and E. Huerta, *Phytochemistry*, 24, 2977 (1985).
- T.F. Stuessy, in: "The Biology and Chemistry of the Compositae, vol. 2." Ed. by V.H. Heywood, J.B. Harborne, and B. Turner, Academic Press, New York, 1977, p. 636.
- E. Rodríguez, G.H.N. Towers, and J.C. Mitchell, *Phytochemistry*, **15**, 1573 (1976).
- M.W. Bierner, Biochem. Syst., 1, 95 (1973).
- 11. M.W. Bierner, Brittonia, 26, 395 (1974).
- H. Minato, S. Nosaka, and I. Horibe, J. Chem. Soc., 5503 (1964).
- W. Herz, C.M. Gast, and P.S. Subramaniam, J. Org. Chem., 33, 2780 (1968).
- P.A. Grieco, Y. Ohfune, and G. Majetich, J. Org. Chem., 48, 360 (1983).
- S. Fortier, G.T. DeTitta, and P.A. Grieco, Acta Cryst., B35, 1742 (1979).
- 16. M. Ul-Haque and C.N. Caughlan, J. Chem. Soc. B, 355 (1967).
- W. Herz, K. Watanabe, and J.F. Blount, J. Org. Chem., 47, 3011 (1982).
- W. Herz, N. Kumar, and J.F. Blount, J. Org. Chem., 46, 1356 (1981).
- R.C. Pettersen and H.L. Kim, J. Chem. Soc., Perkin Trans. 2, 1399 (1976).
- G.M. Sheldrick, "'SHELXTL' revision 3: An Integrated System for Solving, Refining and Displaying Crystal Structures from Diffraction Data," University of Göttingen, Federal Republic of Germany, 1981.
- "International Tables for X-ray Crystallography," vol. 4. Kynoch Press, Birmingham, England, 1974.

Received 11 July 1986

²Atomic coordinates for this structure have been deposited with the Cambridge Crystallographic Data Centre and can be obtained on request from Dr. Olga Kennard, University Chemical Laboratory, Lensfield Road, Cambridge, CB2 1 EW, UK.